

Research

Host specificity and interaction networks of insects feeding on seeds and fruits in tropical rainforests

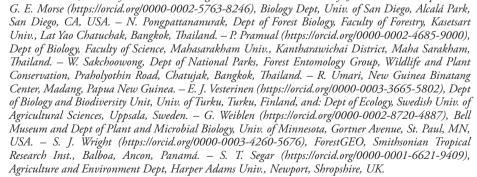
Yves Basset, Leonardo R. Jorge, Philip T. Butterill, Greg P. A. Lamarre, Chris Dahl, Richard Ctvrtecka, Sofia Gripenberg, Owen T. Lewis, Héctor Barrios, John W. Brown, Sarayudh Bunyavejchewin, Buntika A. Butcher, Anthony I. Cognato, Stuart J. Davies, Ondrej Kaman, Petr Klimes, Miloš Knížek, Scott E. Miller, Geoffrey E. Morse, Vojtech Novotny, Nantachai Pongpattananurak, Pairot Pramual, Donald L. J. Quicke, Watana Sakchoowong, Ruma Umari, Eero J. Vesterinen, George Weiblen, S. Joseph Wright and Simon T. Segar

Y. Basset (https://orcid.org/0000-0002-1942-5717) ☑ (bassety@si.edu) and G. P. A. Lamarre (https://orcid.org/0000-0002-7645-985X), ForestGEO, Smithsonian Tropical Research Inst., Apartado, Balboa, Ancon, Panamá, and: Biology Centre of the Czech Academy of Sciences, Inst. of Entomology, Ceske Budejovice, Czech Republic. YB also at: Maestria de Entomologia, Univ. de Panamá, Panama City, Republic of Panama. – L. R. Jorge (https://orcid. org/0000-0003-4518-4328), P. T. Butterill (https://orcid.org/0000-0002-5554-6591), C. Dahl (https://orcid.org/0000-0003-4964-3974), R. Ctvrtecka (https://orcid.org/0000-0002-0599-6850), O. Kaman (https://orcid.org/0000-0003-1480-2904) and V. Novotny (https://orcid.org/0000-0001-7918-8023), Faculty of Science, Univ. of South Bohemia, Ceske Budejovice, Czech Republic, and: Biology Centre of the Czech Academy of Sciences, Inst. of Entomology, Česke Budejovice, Czech Republic. – S. Gripenberg (https://orcid.org/0000-0002-8788-2258), School of Biological Sciences, Univ. of Reading, Reading, Berkshire, UK. - O. T. Lewis (https://orcid.org/0000-0001-7935-6111), Dept of Zoology, Univ. of Oxford, Oxford, UK. - H. Barrios (https:// orcid.org/0000-0002-0542-1731), Maestria de Entomologia, Univ. de Panamá, Panama City, Republic of Panama. – J. W. Brown (https://orcid.org/0000-0001-5610-9855) and S. E. Miller (https://orcid.org/0000-0002-4138-1378), National Museum of Natural History, Smithsonian Inst., Washington, DC, USA. – S. Bunyavejchewin (https://orcid.org/0000-0002-1976-5041), Dept of National Parks, Research Office, Wildlife and Plant Conservation, Bangkok, Thailand. – B. A. Butcher (https://orcid.org/0000-0002-0541-0709) and D. L. J. Quicke (https://orcid.org/0000-0003-4471-6775), Dept of Biology, Faculty of Science, Integrative Ecology Laboratory, Chulalongkorn Univ., Phayathai Road, Pathumwan, Bangkok, Thailand. BAB also at: Center of Excellence in Entomology: Bee Biology, Diversity of Insects and Mites, Chulalongkorn Univ., Pathumwan, Bangkok, Thailand. – A. I. Cognato (https://orcid. org/0000-0001-6436-244), Dept of Entomology, Michigan State Univ., Natural Science Bldg., East Lansing, MI, USA. – S. J. Davies (https://orcid. org/0000-0002-8596-7522), Forest Global Earth Observatory, Smithsonian Tropical Research Inst. and National Museum of Natural History, Washington, USA. – P. Klimes (https://orcid.org/0000-0002-6561-4455), Biology Centre of the Czech Academy of Sciences, Inst. of Entomology, Ceske Budejovice, Czech Republic. – M. Knížek (https://orcid.org/0000-0003-3221-9448), Forestry and Game Management Research Inst., Strnady, Jiloviště, Czech Republic. –

Oikos

130: 1462–1476, 2021 doi: 10.1111/oik.08152

Subject Editor: Anna Traveset Editor-in-Chief: Dries Bonte Accepted 20 May 2021





In the tropics, antagonistic seed predation networks may have different properties than mutualistic pollination and seed dispersal networks, but the former have been considerably less studied. We tested whether the structure of antagonistic tripartite networks composed of host plants, insects developing within seeds and fruits, and their insect

© 2021 Nordic Society Oikos. Published by John Wiley & Sons Ltd

www.oikosjournal.org

parasitoids could be predicted from plant phylogenetic distance and plant traits. We considered subsets of the networks ('subnetworks') at three rainforest locations (Panama, Thailand, Papua New Guinea), based on insect families, plant families or plant functional groups. We recorded 3197 interactions and observed a low percentage of realized interactions, especially in Panama, where insect host specificity was higher than in Thailand or New Guinea. Several factors may explain this, including insect faunal composition, incidence of dry fruits, high fruit production and high occurrence of Fabaceae at the Panamanian site. Host specificity was greater among seed-eaters than pulp-eaters and for insects feeding on dry fruits as opposed to insects feeding on fleshy fruits. Plant species richness within plant families did not influence insect host specificity, but site characteristics may be important in this regard. Most subnetworks were extremely specialized, such as those including Tortricidae and Bruchinae in Panama. Plant phylogenetic distance, plant basal area and plant traits (fruit length, number of seeds per fruit) had important effects on several network statistics in regressions weighted by sampling effort. A path analysis revealed a weak direct influence of plant phylogenetic distance on parasitoid richness, indicating limited support for the 'nasty host hypothesis'. Our study emphasizes the duality between seed dispersal and seed predation networks in the tropics, as key plant species differ and host specificity tends to be low in the former and higher in the latter. This underlines the need to study both types of networks for sound practices of forest regeneration and conservation.

Keywords: Barro Colorado Island, functional group, nasty host hypothesis, plant phylogeny, quantitative food web, seed predation

Introduction

Community assembly and the relationships among interacting species are frequently studied using ecological interaction networks (Jordano et al. 2003, Blüthgen et al. 2006, Poisot et al. 2015, Dáttilo and Rico-Gray 2018), as the structure of these networks may be critically linked to the dynamics and stability of interacting species within the community (Paniagua et al. 2009). Mutualistic networks involving the processes of pollination and seed dispersal have been relatively well studied in tropical rainforests, often emphasizing vertebrates (Olesen and Jordano 2002, Schleuning et al. 2012, Escribano-Avila et al. 2018). To date, antagonistic networks, incorporating information on the frequency of each trophic interaction and emphasizing invertebrates in rainforests have been assembled for leaf miners, gallers, leaf-chewers, their hosts and their parasitoids (review in Morris et al. 2014) and, more rarely, for seed predators (Gripenberg et al. 2019).

In tropical rainforests, insects are the main seed predators, especially before seed dispersal (Janzen 1971). Insects that kill seeds either before or after dispersal influence the population dynamics of individual plant species, and ultimately, plant diversity and assemblage composition (Lewis and Gripenberg 2008). In addition to true seed predators, other functional groups of insects, notably in Diptera and Lepidoptera, feed on the fleshy parts of fruits (Ctvrtecka et al. 2016). This guild of 'pulp eaters' (as opposed to 'seed eaters') can cause fruit abortion and fall, with consequences for plant population dynamics (Stephenson 1981). In the tropics data regarding insect assemblages feeding on seeds/fruits are infrequent (Gripenberg 2018). So far, interaction networks have been built for tephritid flies breeding in tropical flower heads (Prado and Lewinsohn 2004) or in tropical fruits (Novotny et al. 2010), and for the whole assemblage of seed predators in one Panamanian rainforest (Gripenberg et al. 2019).

Concealed insect herbivores, such as seed/fruit predators, are more specialized than insect herbivores that feed externally (Novotny and Basset 2005). Studies in tropical rainforests

have often confirmed the high host specificity of seed/fruit predators (Janzen 1980, Hopkins 1983, Nakagawa et al. 2003, Copeland et al. 2009, Ctvrtecka et al. 2014, Sam et al. 2017, Gripenberg et al. 2019). Since foliar chemistry and plant phylogeny predict patterns of host use by caterpillars in tropical rainforests with high concordance (Segar et al. 2017, Volf et al. 2017), we expect that plant phylogeny may also influence assemblages of seed/fruit insects in tropical rainforests.

Resource availability, such as the production of young leaves, is key to understanding the local distribution of insect folivores in tropical rainforests (Basset 2001). Likewise, it may be crucial to explain the structure and species interactions in assemblages of insects feeding on seeds/fruits in tropical rainforests. In mutualistic networks, biotic specialization decreases with increasing plant diversity, because high plant diversity may reduce relative plant abundance and related plant resources (Schleuning et al. 2012). However, antagonistic and mutualistic networks may be structured differently (Morris et al. 2014).

We expect that plant phylogeny (Segar et al. 2020), local plant diversity and abundance, seed availability and functional plant traits (Basset et al. 2018) may influence interaction networks involving insects breeding in seeds and fruits in tropical rainforests. In this contribution we test whether the structure of antagonistic tripartite networks composed of host plants, insects breeding in seeds and fruits and their insect parasitoids at three representative rainforest locations within different biogeographical regions can be predicted from different plant variables. Because of the high diversity of our study systems, we consider subsets of the overall networks ('subnetworks') at each location, either based on insect families, plant families or plant traits. We answer the following questions:

1) accounting for host-plant phylogenetic relatedness, does herbivore host specificity vary among a) insect families or feeding guilds? b) Plant families or functional groups? And c) three tropical forests? One important variable accounting for network structure is specialization (Blüthgen et al. 2006), which is positively related to host phylogenetic isolation (Jorge et al. 2017). Host specificity is likely to differ among insect guilds associated with fruit pulp versus seeds, as seeds are better chemically protected than pulp (Janzen 1971). Plant traits may also influence seed predator load and host specificity (Janzen 1971, 1980, Basset et al. 2018, Dahl et al. 2019). Low plant richness may favor high insect host specificity, as suggested by comparisons of insect herbivores in temperate and tropical forests (Novotny et al. 2002).

2) Does plant phylogenetic relatedness or plant functional similarity explain the structure of interaction networks between seeds or fruits and their insect predators across study sites or across different local subnetworks?

Related insect herbivores tend to feed on related host plants (Ehrlich and Raven 1964), because related plants may often (Berenbaum 2001, Rønsted et al. 2012) but not always (Sedio et al. 2018) share similar chemical defences. In turn, specialized herbivores, particularly seed predators, may be adapted to detoxify chemical defences (Kergoat et al. 2005). Hence, the structure of plant-herbivore interaction networks may have a strong phylogenetic signal (Weiblen et al. 2006, Segar et al. 2020). Alternatively, plant apparency theory (Feeny 1976) has been incorporated into a framework of three syndromes of plant defence, including: 1) tolerance/ escape; 2) low nutritional quality and; 3) high nutritional quality and defence (Agrawal and Fishbein 2006). Under this framework, plant functional traits may predict the structure of seed predator networks as well as, or better than, plant phylogenetic relatedness.

3) Does the species richness of parasitoid assemblages feeding on seed/fruit predators reflect the traits of host plants, i.e. do the effects of plants on herbivores cascade upwards to affect the next trophic level?

The nasty host hypothesis proposes that insect herbivores feeding on plant hosts with strong and/or distinctive chemical defenses may support reduced loads of parasitoids because herbivore tissues may be more toxic to parasitoids (Gauld et al. 1992). Thus, this hypothesis predicts that plant phylogenetic relatedness, as a surrogate for plant chemistry (Berenbaum 2001), a) should have a significant effect on the species richness of parasitoids; b) that this effect should be strong, and; c) that this effect should also be positive (i.e. mean phylogenetic distance is predicted to be negatively correlated with parasitoid species richness).

Material and methods

Study sites

Our study sites are three ForestGEO lowland rainforest plots (Anderson-Teixeira et al. 2014) located in different biogeographical regions. Salient characteristics of the plots and

seed/fruit samples are detailed in Basset et al. (2018) and in Table 1. Neotropical: Barro Colorado Island (BCI) is a 1500-ha island created by the opening of the Panama Canal in 1914. The 50-ha plot is in the heart of the island, which is near the center of more than 700 km² of protected forests. Oriental: the 24-ha plot at Khao Chong (KHC) is in the protected forest of the Khao Ban Thad Wildlife Sanctuary in southern Thailand. Australasian: the 50-ha plot is located in the 10 000 ha Wanang (WAN) Conservation Area in Papua New Guinea. Marked differences between BCI and the two other sites include lower plant richness, higher percentage of species with dry seeds, higher percentage of Fabaceae species, higher average seed rain per plant species and lower ratio of realized to potential interactions (Table 1).

Plant surveys, phylogeny and functional traits

Field methods were similar for all study sites (details in Basset et al. 2018, Gripenberg et al. 2019). Plant surveys spanned several years at each site (Table 1, Supporting information). During the first study year at each site, we indiscriminately surveyed seeds and fruits of locally abundant tree, shrub and liana (more rarely herb) species, to obtain an overview of the local community. During subsequent study years at KHC and WAN, we restricted our sampling effort to the 10 plant families that were most common at each plot. Eight of these focal families were common to all sites: Annonaceae, Arecaceae, Euphorbiaceae, Fabaceae, Lauraceae, Meliaceae, Rubiaceae and Sapindaceae. Unless specified, results are detailed for all host plant species. Seeds and fruits collected on plants or freshly fallen (without apparent decomposition) were opportunistically surveyed within and/or near permanent plots (from an area < 1500-ha corresponding to the smallest study area, BCI). Rearing sample units consisted of 1-200 seeds and/or fruits collected from a single plant. We targeted as many individual plants as possible for each species, typically > 5. To evaluate the phylogenetic relationships between sampled host plant species at each site, we estimated the relationships between our focal species using the R package S.PhyloMaker (Qian and Jin 2016). We used the updated phylogeny and node ages derived from a sequencebased study by Zanne et al. (2014) as a Megatree. Our focal species were placed within the Megatree where possible and placed to family where not possible. This procedure generates three alternative topologies which differ with respect to the resolution of unplaced taxa. We selected the phylogeny derived from 'Scenario 3' as this has been shown to be robust to uncertainty at the higher taxonomic level (Qian and Jin 2016). Note that polytomies in the phylogeny underestimated DSI* slightly for herbivores feeding on a few species within a family. However, we expect this effect to be quite small, especially because it does not apply to monophages that would still have maximum specialization.

To obtain similarity matrices of plant functional traits for each site, we first compiled a matrix of functional traits relevant to seeds and fruits for each plant species, including numerical and categorical variables (Supporting information).

Table 1. Salient characteristics of the study sites and their plant, insect and network interactions. Plot data are from Anderson-Teixeira et al. (2014).

Variable	BCI	KHC	WAN	
Site physiognomy and structure				
Coordinates	9.15°N, 79.85°W	7.54°N, 99.80°E	5.24°S, 145.08°E	
Elevation (m)	120–160	120–330	90–180	
Annual average rainfall (mm)	2551	2665	3366	
Annual average daily maximum air temperature (°C)	26.3	27.1	26.5	
Number of tree species/genera/families recorded in plot	299/181/59	593/285/82	508/245/77	
Percentage of plant spp. with dry seeds/fruits	56.8	26.0	28.0	
Percentage of Fabaceae species to total spp. richness in plot	14.0	3.1	4.8	
Total fruit production (seed rain; dry $g \times m^{-2} \times year^{-1}$)	108.0	7.0	10.8	
Average fruit production per species (dry g \times m ⁻² \times year ⁻¹)*	0.596	0.141	0.157	
Plant samples				
Years of collection	2010–2013	2013–2015	2013–2015	
Number of plant species/plant families surveyed	497/82	357/66	332/67	
No. plant species surveyed within the 10 focal families	218	171	170	
Total number of seeds or fruits collected	208 508	39 252	122 976	
Total weight of samples (kg)	380.2	343.2	439.9	
Insect samples				
Total number of insects reared	27 610	17 555	35 434	
Number of individuals/species of seed eaters**	11 059/311	2100/59	3935/77	
Number of individuals/species of pulp eaters**	5670/214	7265/161	9403/193	
Number of individuals/species of parasitoids***	775/161	359/61	961/96	
Interactions				
Number of interactions realized/% realized- potential****	892/0.26	917/1.01	1388/1.20	
Plant species with most seeds/fruit reared	Mikania leiostachya	Caryota mitis	Mastixiodendron pachyclados	
Most abundant herbivore species	Pagiocerus frontalis	Coccotrypes myristicaceae	Coccotrypes sp.n.3	
Most abundant parasitoid species	Dorylinae sp. 156	Alysiinae sp. 13	Diospilus sp. 2	
Network statistics for full network (level 1–2)				
Average DSI* ± SE	0.906 ± 0.013	0.577 ± 0.029	0.503 ± 0.028	
Number of compartments	85	20	9	
H2'	0.914	0.664	0.657	
Generality	1.85	6.84	5.15	
Nestedness	0.96	1.61	2.20	
Vulnerability	2.23	4.69	6.05	
Connectance	0.006	0.021	0.022	

^{*}Plant species recorded in litterfall traps.

We then used hierarchical daisy clustering methods to identify functional groups. Finally, we used a mixed principal component analysis (PCA) for numerical and categorical variables to interpret the functional groups. Scores of each plant species on the PCA axes were used to build similarity matrices of functional traits that were used in subsequent analyses. In sum, 1186 plant species could be assigned to one of five functional groups, coded A, B, C, D or E (A: often fleshy green fruits; B; often dry dehiscent fruits; C: often fleshy orange fruits; D: often red fruits; E: often small fleshly black-green fruits). The Supporting information provides details about the composition of the matrix of functional traits, computational steps and relevant references, as well as the results obtained.

Insect rearing and processing

Methods for rearing seed/fruits insects are detailed in Basset et al. (2018) and the Supporting information. Insects were identified with the assistance of taxonomists (Basset et al. 2018) and/or by molecular techniques (Supporting information). Insects reared from seeds/fruits were assigned to the following guild categories (Basset et al. 2018): seed eaters (coded as SE: larva feeding mostly on seed tissue), pulp eaters (PU: larva feeding mostly on mesocarp tissue) and parasitoids (PA: larva feeding on insect hosts). Seed and pulp eaters consisted mainly of seven taxa that represented most of the material reared and are considered in analyses restricted

^{**}Seven focal taxa only, 'Methods' section.

^{***}Braconidae and Ichneumonidae only.

^{****}No. of interactions between levels 1-2 and 2-3; percentage of realized to potential interactions.

to insect taxa: Bruchinae, Scolytinae, Curculionidae others than Scolytinae (Coleoptera), Tortricidae, Pyralidae (Lepidoptera), Stratiomyidae and Tephritidae (Diptera). Bruchinae were well represented only at BCI. Hereafter for sake of simplicity, we refer to these seven taxa as 'insect families'. For parasitoids, analyses were restricted to Braconidae and Ichneumonidae because they represented most (69%) of the parasitoids reared from samples and their taxonomy was supported by molecular data.

Interactions, topologies and subnetworks

Trophic relationships were inferred from the number of primary consumers reared from samples of seeds/fruits of host plant species at the first trophic level (coded as level 1–2). For the third trophic level, we considered interactions between Braconidae and Ichneumonidae and their insect hosts (coded as level 2-3). Contrary to interactions between the first and second trophic levels, third level data only reflected expected interactions, not documented interactions, because parasitized hosts were not isolated and reared individually, the parasitoids instead being reared from samples including relatively high numbers of seeds and fruits. To assign putative hosts to each parasitoid species, we applied three simple rules, as detailed in the Supporting information: 1) since many parasitoid lineages are rather conservative in host use, we followed Quicke (2015) to select the most likely host order or family; 2) in case of conflicts, we examined for each parasitoid species the consistency of co-occurrence with the putative host species in all samples from which the parasitoid species was reared; 3) eventually, we considered the highest abundance of putative host reared in samples in which the parasitoid species was also reared. We considered expected interactions between hosts plants and parasitoids (coded as level 1-3) to answer Ouestion 3.

We constructed tripartite and quantitative interaction networks for the three full networks at BCI, KHC and WAN and for meaningful subsets of the data. This approach was selected because of the complexity of the full networks, which involved an order of magnitude more interacting species than most published networks (Schleuning et al. 2012, de Aguiar et al. 2019). Breaking a complex network into smaller sub-networks can reveal interesting patterns (Lewinsohn et al. 2006, de Aguiar et al. 2019) and has been performed with networks including insects (Quinto et al. 2012), which are far more diverse than those based on vertebrates (Schleuning et al. 2012). Each of our three full networks can be viewed as a collection of empirical subnetworks built by sampling interactions of a particular taxonomic/ functional group within a locality, a general approach which is consistent with most published networks (de Aguiar et al. 2019). In addition, subnetworks were relatively independent from each other, thus motivating analyses at the level of subnetworks ('subnetwork structure' below). The meaningful subsets (hereafter 'topologies' for sake of brevity and in reference to how subnetworks are arranged) were based on; A) insect taxa: the distribution of particular insect families

on plant species (n = 7 taxa, resulting in 19 subnetworks); B) plant family: the distribution of insect species within particular plant families (n = 8, 24 subnetworks); or C) plant functional groups: the distribution of insect species on particular plant functional groups (n=5, 15 subnetworks). Topology (A) is more relevant to questions 1 and 3 of the Introduction, whereas topologies (B) and (C) are more relevant to question 2. Topologies (A) and (C) included insects reared from all host plants at each site, whereas for topology (B) we restricted the data to focal plant families. Further, for topologies (B) and (C) we also included in subnetwork illustrations plant species that were surveyed but yielded no reared insects (these 'empty hosts' were not considered in the calculation of subnetwork statistics, below). Variables as surrogates of either resource availability or sampling effort are discussed in Basset et al. (2018). Here, we consider that resource availability is most accurately tracked as the square root of the number of seeds collected for each plant species.

Data analyses

Question 1. Herbivore host specificity

Quantitative metrics accounting for network-wide specialization (Blüthgen et al. 2006, Dormann et al. 2009) may be biased by sample size (Morris et al. 2014) and by nonrandom sampling of the plant phylogeny (Redmond et al. 2018). To overcome these challenges, we calculated herbivore specificity with the rescaled distance-based specialization index (DSI* - Jorge et al. 2014, 2017), which measures trophic specialization by accounting for host phylogenetic relatedness and resource availability. This quantitative metric accounts for differences in abundance and sampling effort of consumers and is largely independent of sample size. Briefly, DSI* measures specialization as a deviation from a random expectation involving the mean pairwise phylogenetic distance between hosts and is rescaled to enable the comparison of consumers that differ in their recorded sample sizes. DSI* varies between -1 (maximum achievable generalization) and 1 (monophages or maximum achievable specialization; Jorge et al. 2017). At each site we calculated DSI* for all seedand pulp-eating insect species considering all host plant data available. DSI* was not calculated for parasitoid species as, due to many missing data, we could not build reliably a phylogeny for insect herbivores.

To answer questions 1A–1C, we first tested for differences in DSI* among herbivore taxa and guilds (pulp and seed eaters), plant families and functional groups, and study sites. We used non-parametric Kruskal–Wallis tests because DSI* values were skewed towards high specialization. Then, we used DSI* as the response variable in a model including taxa, guilds, sites and taxa:site and guild:site interactions as independent variables. To control for variation among insect families on DSI* (excluding Bruchinae, collected only at BCI), we performed a linear mixed model with site and insect guild as fixed effects and insect family as a random effect. To control for variation among plant families, we performed a linear mixed model with mean DSI* of the herbivores feeding on

each plant species as the dependent variable, site and plant functional groups as fixed factors, and plant family as a random factor. To evaluate the effect of local plant richness, we calculated the correlation between the average DSI* of the insect assemblage feeding on each focal plant family at each study site (n=24 site-family combinations) and the species richness of these plant families at each site.

Question 2. Subnetwork structure

We addressed question 2 by modeling indices for network properties at the level of the full network or subnetwork (hereafter 'network statistics'). Rather than focusing on any single metric, we calculated the following standard network statistics reflecting network structure (Morris et al. 2014): degree of compartmentalization (number of compartments); weighted quantitative network specialization index H2'; weighted quantitative generality (effective number of host species per consumer species); nestedness (specialization asymmetry); weighted quantitative vulnerability (effective number of consumer species per host plant species); and connectance (degree of redundancy in the study system). The Supporting information describes these network statistics in greater detail. All network statistics were calculated with the R package Bipartite (Dormann et al. 2018). We also reported the number of species in both trophic levels and the sum of links for each subnetwork, as well as sampling intensity sensu Schleuning et al. (2012). Models were estimated separately for topologies A-C (insect families, plant families and plant functional groups) and trophic levels 1–2. We considered network statistics as independent data points in models, because of: a) no overlap between insect and plant species across study sites; and b) for a particular site and topology, the average pairwise species overlap between subnetworks was 4.3%, 2.2% and 4.1% for herbivore families, plant families and functional groups, respectively. Network statistics were also calculated for trophic levels 2-3 and 1-3 (question 3 below). We did not calculate network statistics when subnetworks were too small (number of species in the lower level < 5).

We used null models to assess how network metrics deviated from those expected from a random distribution of interactions. Null models were implemented for three full networks and 58 subnetworks using Patefield's algorithm ('r2dtable' in Bipartite's nullmodel function, Dormann et al. 2018), in which the marginal species totals are constrained as per the respective observed networks. We ran 1000 randomisations for subnetworks and 200 for full networks (due to the time and CPU demands of running analyses on large sparse networks). We evaluated whether network statistics differed significantly between study sites by performing simple Kruskal-Wallis and Dwass-Steel tests. We refined this analysis by using three types of regression models. To account for the effect of sampling effort, the number of observed interaction events (i.e. the number of links; Schleuning et al. 2012) was used as a weighting factor in each regression. The first type of regression (hereafter 'model type I') included mixed models with network statistics as dependent variables, sites

and insect guilds as fixed factors and topologies (insect and plant families, plant functional groups) as random factors.

Following Chamberlain et al. (2014), we modeled network statistics with beta regression (H2' and connectance), generalized linear models with Poisson (number of compartments) or Gaussian (all other statistics) error distribution, separately for each topology A–C. Models type II parsed the effects of plant phylogeny and of plant ecological variables (resource and functional traits), and were calculated as:

Network statistic = MPD + FDis

where MPD is the average plant relatedness (mean phylogenetic distance between plant species included in subnetwork, calculated with the function mpd of the R package Picante, Kembel et al. 2010), and FDis is the functional dispersion within the subnetwork, calculated with the function fdisp of the R package FD (Laliberté et al. 2014). FDis quantifies trait diversity as the mean distance in multidimensional trait space of individual plant species to the centroid of all species (Laliberté and Legendre 2010). FDis was calculated with variables accounting for: a) plant resource (no. of stems and basal area in ForestGEO plots, seed rain (g dry weight \times m⁻²), equivalent to total fruit biomass and estimated from litterfall traps, Basset et al. 2018); and b) plant traits (fruit length and weight (partly related to seed size and biomass), number of seeds per fruit). We used the function betareg of the R package betareg to perform beta regressions (Gruen et al. 2012). For other regressions, we performed model simplification to extract the variables with significant predictive power with the built-in functions glm and step (backward selection of variables) of the R package (<www.r-project.org>). We eventually tested the significance of estimators by an ANOVA (type 2 test) with the function anova in the R package 'car' (Fox and Weisberg 2019).

A last series of models considered more specifically the effects of plant variables (hereafter 'models type III'):

Network statistic = Plant species richness + MPD + CWM₁
+ CWM₂ + ... +
$$CWM_n$$

where Plant species richness was the number of confamilial species in ForestGEO plots (for topology B) or the number of plant species in functional groups in ForestGEO plots (for topology C), and CWM_n was the community weighted mean of trait n, weighted by the number of samples collected and calculated with function dbFD of R package FD (Laliberté et al. 2014; plant species richness could not be included as an independent variable for the topology based on herbivore families). Independent variables accounted for sampling effort, plant species richness, mean phylogenetic distance, plant resource and plant traits (as defined in models type II; only continuous variables). Before analyses, highly correlated variables (r > 0.7) were removed from models.

Regressions were calculated and the significance of estimators tested as described previously.

Question 3. Parasitoid species richness

To approach question 3, we computed a path analysis with a bottom-up flow of correlations implying direct and indirect correlations between herbivore species richness, parasitoid species richness (dependent variables) and selected independent variables. This analysis was performed at the level of the plant species, considering all plant species (n=618)at the three sites from which seed predators were reared. Independent variables included mean phylogenetic distance and variables related to plant resource or plant traits (Supporting information). They were selected based on a) a rationale for each path explained in Supporting information; and b) the best predictors in the regressions performed previously (previous section and Results). The mean phylogenetic distance of a plant species to all other plant species was calculated with the function cophenetic of the R package Picante (Kembel et al. 2010). The model was calculated with the Ω nyx software (von Oertzen et al. 2015).

Data deposition

Interaction data were deposited in Figshare https://doi.org/10.25573/data.11444571.v1. Molecular insect data were deposited in the following Barcode of Life projects (BOLD, www.boldsystems.org): BCI: 2310 sequences in projects BCISP and PSPLP; KHC: 398 sequences in KHCSP and KHCTE; WAN: 1646 sequences in WANSP, FRUT and CURCU. Full data for specimens sequenced (including those that failed), including images and host plants, are available on BOLD, accessible by DOI for the datasets:

dx.doi.org/10.5883/DS-BCISP (BCI), dx.doi.org/10.5883/DS-KHCFRUIT (KHC) and dx.doi.org/10.5883/DS-PNGFRUIT (WAN).

Results

We collected 1163 kg of seeds and fruits, which produced 80 600 insects across the three sites (Table 1). The composition and species richness of the insect material is discussed elsewhere (Basset et al. 2018). This contribution analyzes the 3197 interactions across a total of 1176 plant, 1015 herbivore and 318 parasitoid species at the three study sites (Table 1). Only 0.58% of the potential 553 160 interactions were realized (Table 1). Since most properties of subnetworks do not represent properties of whole networks (Jordano 2016), we detail network statistics for the full networks (level 1–2) of BCI, KHC and WAN in Table 1, for comparison with other studies. These results emphasize differences between BCI and the other two sites, which we analyze in more depth by considering subnetwork data.

Question 1. Insect host specificity

Differences in the median value of DSI* across insect families were significant (W=114.0, p < 0.001, df =6; Fig. 1b). Bruchinae were by far the most specialized taxa, followed by Pyralidae, Curculionidae, Scolytinae, Tortricidae, Tephritidae and Stratiomyidae (Fig. 1b). Seed eaters were significantly more specialized than pulp eaters (Mann–Whitney test, U=91.8, p < 0.001, df=1; Fig. 1c). There were also significant differences between the median DSI* of insect faunas feeding across plant families (W=50.9, p > 0.001, df=7). For example, on average, insects feeding on Fabaceae were

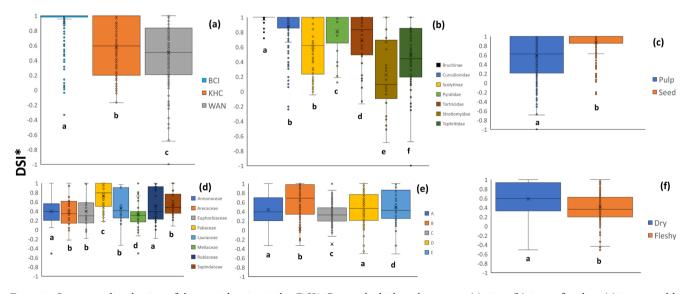


Figure 1. Summary distribution of the specialization index DSI*. Box and whisker plots across (a) sites, (b) insect families, (c) insect guilds, (d) plant families, (e) seed functional groups and (f) categories of fruit fleshiness. Groups with different letters are significantly different (Dwass–Steel tests, p < 0.05).

rather specialized whereas those feeding on Meliaceae were less so (Fig. 1d). Differences in median of DSI* for insects feeding across plant functional groups were also significantly different (W=53.1, p < 0.001, df=4). In particular, insects feeding on plants belonging to functional group B (dry dehiscent fruits) were far more specialized than those feeding on group C plants (fleshy orange fruits; Fig. 1e). Overall, insects feeding on dry fruits were significantly more specialized than those feeding on fleshy fruits (Mann–Whitney test, U=39.9, p < 0.001, df=1; Fig. 1f). The percentage of true monophagous species (DSI*=1) was higher at BCI (69.5%) than at KHC (25.3%) and WAN (18.9%) and the median of DSI* was significantly different across sites (Kruskal–Wallis test, W=201.8, p < 0.0001, df=2; Fig. 1a).

The mixed linear model with DSI* as dependent variable and insect family as random factor indicated that the effect of site was significant, but not that of insect guild (seed eater versus pulp eater; Supporting information). When considering the mean DSI* of insect species feeding on plant families, a similar model indicated that the effects of both site and plant functional group (coded A–E, Supporting information) were significant, but not their interaction (Supporting information). There was no correlation between the average DSI* of insects feeding on the eight focal plant families at the three study sites and the local species richness of these plant families (r=-0.16, p=0.45; Supporting information).

Question 2. Variables affecting the structure of subnetworks

We illustrate nine of the 58 subnetworks studied (Fig. 2) and detail characteristics of all subnetworks in the Supporting information. In null models, most (95.6%) observed network statistics deviated significantly from those expected from a random distribution of interactions (Supporting information), with nestedness involved in nearly all cases where the deviation was not significant (Supporting information). The distribution of the six main network statistics is summarized in Fig. 3 for the topology based on insect herbivore families and trophic levels 1-2. BCI subnetworks had significantly more compartments, significantly higher degree of specialization (H2'), and significantly lower effective number of host species per consumer species (generality) than KHC and WAN. In particular, for all herbivore families, H2' was also higher at BCI than at other sites (Supporting information). Subnetworks based on Bruchinae, Tortricidae, Curculionidae and Pyralidae were in general more specialized than those based on Tephritidae, Scolytinae and Stratiomyidae (Supporting information). Food webs based on stratiomyid flies were rather unspecialized, rarely parasitized by braconids and their subnetwork at WAN represented the most unspecialized subnetwork of all subnetworks analyzed. Conversely, the most specialized subnetwork was based on Tortricidae at BCI, followed closely by Bruchinae at BCI (Supporting information). Other network variables were not significantly different between sites (Fig. 3). Mixed models weighting the effect of sampling effort (models type I), for the topology based on insect families, confirmed the strong effect of sites on the number of compartments, H2' and generality (Supporting information).

For the topology based on plant families, BCI had significantly more compartments than KHC and WAN, and KHC had significantly larger effective number of consumer species per host plant species (vulnerability) than BCI and WAN (Supporting information). Nestedness and Connectance were not significantly different between sites for any of the topologies considered (Supporting information). For the topology based on plant functional groups, the number of compartments and H2' were significantly higher at BCI than at the other sites, whereas generality and vulnerability were significantly lower at BCI (Supporting information). Mixed models weighting the effect of sampling effort (models type I) for both topologies based on plant families and functional groups confirmed the significant effect of site on all network statistics (Supporting information).

When parsing the effects of plant phylogeny and ecological variables (models type II, Supporting information), the significance of effects could be ranked overall as mean phylogenetic distance (MPD) > functional dispersion (FDis). MPD was a significant predictor of network statistics calculated for plant functional groups (topology C), while FDis was more important to predict network statistics calculated for plant families (topology B). Note that for models based on plant families, the effect of MPD may be low due to the limited range of MPD within plant families. This also applies for FDIs in models based on plant functional groups.

Not surprisingly, models best explained by CWM (models type III) were related to plant functional groups (65–98% of variance explained, Table 2). Over the different topologies, the variation explained by the type III models was greatest for number of compartments and least for generality. Several variables were reasonably good predictors of subnetwork structure, in order of importance seed rain, mean phylogenetic distance and number of plant species, as well as number of seeds per fruit (Table 2). Plant species richness was a good predictor of network statistics (connectance, nestedness, generality), only for topology B based on plant families. In models describing network statistics for plant functional groups, variables related to plant traits were important to predict H2', while variables related to plant resources were important for number of compartments, vulnerability, connectance and nestedness (Table 2).

Question 3. Species richness of herbivores and parasitoids

Our path analysis model explained 19% of the variance in the number of parasitoid species supported by each plant species (Fig. 4). As expected, significant paths existed between plant traits and herbivore species richness, and between plant resources and herbivore species richness. Mean plant phylogenetic distance influenced plant traits but not directly herbivore species richness. The strongest direct paths (as judged from standardized path coefficients) influencing parasitoid

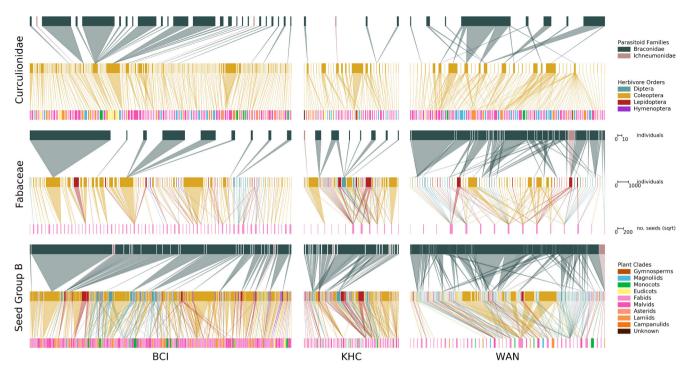


Figure 2. Examples of interaction subnetworks at BCI (left), KHC (middle) and WAN (right). Top: topology based on insect family, here the Curculionidae (without Scolytinae). Middle: topology based on plant family, here the Fabaceae (including plant species lacking insects attacking seeds or fruits). Bottom: topology based on plant functional group, here Group B (dry fruits, protected, dehiscent and relatively large, Supporting information). For each subnetwork, the abundance of parasitoid species (top series of rectangles) and herbivore species (middle series of rectangles) are represented by the number of individual reared, whereas the abundance of plant species (bottom series of rectangles, coloured by plant clades following APG IV: Chase et al. 2016) are represented by the square root of the number of seeds collected. Parasitoid families, herbivore orders and plant clades are identified by distinct colours as coded on the right. The scale for each level is also indicated on the right. All subnetwork nodes are ordered as to minimize the number of crossed interactions. From left to right and top to bottom these subnetworks are coded as HB-CURC-BCI, HB-CURC-KHC, HB-CURC-WAN, PL-FABA-BCI, PL-FABA-KHC, PL-FABA-WAN, FG-B-BCI, FG-B-KHC and FG-B-WAN in the Supporting information.

species richness originated from herbivore species richness (positive), mean phylogenetic distance (negative) and basal area (positive). Thus, although the effect of mean plant phylogenetic distance was significant and negative on parasitoid species richness (as predicted by the nasty host hypothesis of Gauld et al. 1992), its direct path was about five times smaller than the corresponding direct path originating from herbivore species richness, pointing to other explanations.

Discussion

In this contribution we examined the interaction networks involving seeds and fruits, the insects feeding on them and their parasitoids, at three tropical sites. To analyze the 3197 interactions reported, we considered three 'topologies' (how subnetworks are arranged) resulting in 58 different subnetworks, which were largely independent from each other. This strategy was possible because of the very low overlap of interacting species between subnetworks but may not be applicable to other types of networks, such as mutualistic networks. Topology A, based on families of seed predators, may be useful to entomologists, whereas topologies B and C (based on plant families and functional groups) may be more interesting to

botanists. Some interactions may not have been documented in our study system, since attack rates were rather low (8.5% of seeds/fruits attacked, Basset et al. 2018) and substantial sampling effort may be required to rear insects attacking seeds and fruits, For example, Ctvrtecka et al. (2014) consider a minimum sample size of 5 kg of fruits/seeds per plant species adequate to rear weevils feeding on fruits/seeds. This condition was achieved for only 3% of our plant species. Low sampling effort may result in inflated insect host specificity and network specialization (Blüthgen et al. 2006). Sampling effort in the field (collecting seeds/fruits, rearing insects) was higher at BCI than at the other sites (Table 1). Hence, we believe that the high host specificity documented at BCI is not an artefact. Another obvious limitation in our study was the indirect documentation of linkages between insect herbivores and parasitoids. Some of the linkages reported here will need confirmation but given the limited data on tropical seed predators they are nevertheless valuable.

Insect host specificity

Insect host specificity varied significantly between insect families. While some seed- or fruit-feeding taxa are known to be extremely specialized in rainforests (Bruchinae: Janzen

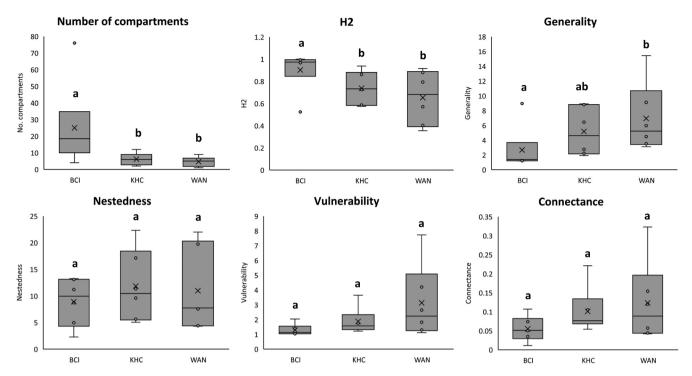


Figure 3. Summary distribution of the six main network level statistics across study sites (BCI, KHC, WAN) for subnetworks based on insect herbivore families and trophic levels 1-2 (plants–insect herbivores; n=18). The Bruchinae subnetwork for BCI was not included as it has no equivalent at other study sites. Groups with different letters are significantly different (Dwass–Steel tests, p < 0.05).

1980, Curculionidae: Ctvrtecka et al. 2014), others are less so (Tephritidae: Novotny et al. 2010). These trends were confirmed in our study, which also indicated that Stratiomyidae, a taxon rarely considered in studies of frugivorous insects, are less specialized pulp eaters than Tephritidae. Seed-eaters were more host-specific than pulp-eaters, confirming that insect host specificity for tropical herbivore guilds in both temperate and tropical forests decreases in the sequence: seed-eaters > leaf-miners > pulp-feeders > leaf-chewers > sap-suckers > xylophages > root-feeders (Novotny and Basset 2005). This partially reflects the plant's allocation of nitrogen and chemical defences to the tissues consumed by these guilds, as young leaves are sometimes better defended than seeds (Janzen 1971, 1980, Bazzaz et al. 1987, Zangerl and Bazzaz 1992, Kergoat et al. 2005). Insects feeding on dry fruits were also more host specific than those feeding on fleshy fruits. Insect host specificity varied significantly among plant families and functional groups, and the effect of site was important in most of our analyses. Overall, we observed the lowest percentage of realized interactions and highest insect host specificity at BCI. This trend was apparent when considering both entire networks (Table 1) and subnetworks (Fig. 1, 3). Similar levels of host specificity for entire networks and across subnetworks may be explained by the preponderance of highly specialized fruit/seed consumers, with very few generalist consumers present in more than one subnetwork.

This high insect host specificity at BCI is likely to result from the following factors: 1) insect faunal composition: highly host specific and diverse Bruchinae are prevalent at BCI and absent from KHC and WAN; 2) fruit fleshiness: BCI has the highest proportion of dry fruits (supporting highly host specific insects), possibly because of lower rainfall at BCI as compared to our other sites (Kissling et al. 2009); 3) fruit production: BCI has on average four times higher seed rain per plant species than KHC or WAN (i.e. high fruit production and low plant species richness: Table 1). Our regressions confirmed the importance of variables such as basal area or seed rain to predict insect specialization; 4) Fabaceae: there is a high percentage of Fabaceae species at BCI (Table 1), which support many host-specific insect species.

In mutualistic networks, such as pollination and seed dispersal networks, specialization decreases with increasing plant diversity. An explanation may be that high plant diversity reduces relative plant abundance and related plant resources, resulting in hard evolutionary constraints on specialists (Schleuning et al. 2012, Escribano-Avila et al. 2018). Antagonistic networks may be different (Morris et al. 2014). Lewinsohn and Roslin (2008) discuss the species richness and host specificity of folivorous insects in tropical rainforests, and our BCI data appear to follow their contention that high species richness may be promoted by more insect species per plant species (Table 1) or higher herbivore host specificity (Fig. 1a). In sum, low plant richness and high insect host specificity at BCI suggests that antagonistic networks based on seed predation may follow the same rules as mutualistic networks, with low plant richness strengthening interactions and favoring high insect host specificity (Novotny et al. 2002).

Table 2. Summary of the best regression models (type III) with network statistics as dependent variables, for each topology considered (A–C). Independent variables include mean phylogenetic distance (MPD) and the community weighted mean (CWM) of functional traits (text for details).

Topology/subnetwork variable	Significant variables	R^2	F	р	AIC
(A) Insect families					
Specialization H2'	MPD***, Fruit length***, Basal area***, No. seed per fruit***, Seed rain***	0.400	2457.1	< 0.001	-3670.1
Connectance	n.s.	_	_	_	_
No. compartments	Seed rain***, Fruit length***, MPD***, Basal area***, No. seed per fruit***	0.581	2637.9	< 0.001	26903.0
Generality	MPD**	0.500	15.9	< 0.001	104.1
Nestedness	n.s.	_	_	_	_
Vulnerability (B) Plant families	Fruit length*	0.342	5.2	0.022	94.5
Specialization H2'	No. seed per fruit***, Basal area***, MPD***, Fruit length***, Seed rain***	0.107	446.1	< 0.001	-1999.3
Connectance	No. plant species***, MPD***, Basal area***, No. seed per fruit***, Fruit length***	0.618	1948.5	< 0.001	-7143.4
No. compartments	Basal area***, Seed rain***, Fruit length***, No. plant species***, MPD**	0.925	18.5	< 0.001	5481.6
Generality	No. plant species*, Fruit length*	0.351	3.4	0.037	44.6
Nestedness	No. plant species***	0.467	8.8	0.002	150.4
Vulnerability	n.s.	_	_	_	_
(C) Plant functional groups					
Specialization H2'	No. seeds per fruit***, Basal area***, Seed rain***, MPD***, Fruit length***	0.696	4169.4	< 0.001	-10018.1
Connectance	Seed rain***, MPD***, Fruit length***, No. seed per fruit***, Basal area***	0.749	5125.6	< 0.001	-20987.9
No. compartments	Seed rain*** , Basal area***, Fruit length***, No. seed per fruit***, No. plant species***	0.987	56.9	< 0.001	13211.0
Generality	n.s.	_	_	_	_
Nestedness	MPD***, Basal area*	0.650	11.2	0.002	60.7
Vulnerability	Seed rain***, Basal area**, MPD*	0.835	9.1	0.002	52.3

^{***}p < 0.001.

Independent variables listed according to decreasing standardized coefficients, best estimators indicated in bold.

Subnetwork structure

Our analyses emphasized the strong effects of site on the different network statistics. Plant assemblages at different rainforests may be phylogenetically different or may possess different traits, or both. Tree assemblages are phylogenetically distinct in many rainforests (Webb 2000) and there are important differences in seed functional traits between our rainforest sites (Supporting information; Dahl et al. 2019). Plant phylogenetic distance had an important effect on subnetwork structure (e.g. specialization H2' and generality in subnetworks based on insect families), but this effect was not overwhelming, as variables related to plant traits or plant resource were also important in this regard. A more explicit inclusion of the hierarchical structure of phylogenies in predicting interaction identities might provide increased explanatory power. Ideally, further analyses would include phylogenies for hosts, herbivores and parasitoids (Ives and Godfray 2006), but herbivore and parasitoid phylogenies are not currently available. Plant traits such as fruit length and number of seeds per fruit, were important predictors of network statistics (Table 2). Other variables related to host phenology, such as the duration of fruiting season and its

synchronization within/among years, may well be important in this regard (Janzen 1976), but they could not be tested in this study, for lack of reliable data at all sites. Variables accounting for plant resource (basal area, seed rain) were also important whereas the effect of mean phylogenetic distance was not excessive. This would lend support to the modified plant defence theory (Agrawal and Fishbein 2006). Both plant resource and plant traits were reasonably good predictors of subnetwork structure, particularly for models based on plant functional groups, emphasizing the interest in this topology as a predictive framework for subnetwork structure.

Upward cascades in the subnetworks

The nasty host hypothesis (Gauld et al. 1992) argues that tropical plants often possess highly active chemical defenses, which may lead to greater host specialization and sequestration of secondary compounds in insect herbivores, and reduced loads of parasitoids on particularly well-defended host plants. However, to date, evidence in favor of this hypothesis is mixed (Quicke 2012, Morris et al. 2014). Alternatively, Smilanich et al. (2009) observed that secondary metabolites sequestered by herbivores may compromise their immune

^{**}p < 0.01.

^{*}p < 0.05.

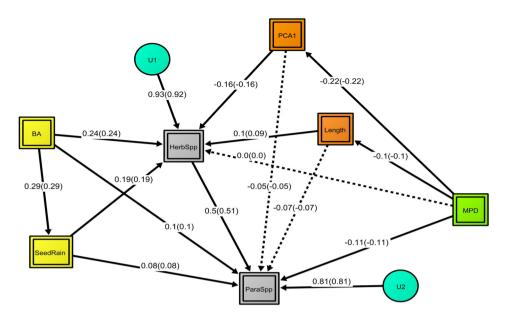


Figure 4. Results of path analysis testing direct and indirect correlations beween the species richness of seed- and fruit-eating insects (HerbSpp), parasitoid species richness (ParaSpp), mean phylogenetic distance (MPD), plant traits (fruit length, length and ordination scores delineating plant functional groups, PCA1) and plant resource (basal area, BA and seed rain, SeedRain), for 618 host plant species. Standardized path coefficients are in parentheses. Significant (p < 0.05) and insignificant paths are indicated by solid and dashed lines, respectively. The rationale of the model is detailed in the Supporting information.

response, making them more vulnerable to successful parasitism (the 'vulnerable host hypothesis'). In our study system, we used plant phylogenic distance as a surrogate for plant chemistry (Berenbaum 2001), as chemical data for tropical fruits and seeds are limited (Gripenberg et al. 2018). Our path model indicated that most of the explained variance in parasitoid species richness on host plants could be attributed to a direct path originating from herbivore species richness, whereas the corresponding path originating from mean plant phylogenetic distance was less important.

The nasty host hypothesis (Gauld et al. 1992) explains parasitoid loads on plants principally with regard to plant chemistry. Were this hypothesis correct, we would have expected a large direct path from mean plant phylogenetic distance to parasitoid species richness. The larger direct path observed from herbivore species richness to parasitoid species richness seems rather consistent with both the resource concentration and resource base hypotheses (Root 1973, Price 1992), predicting that local assemblages of parasitoids may be more diverse when their herbivore hosts are diverse (Hawkins and Lawton 1987) and vulnerable (Smilanich et al. 2009).

Conclusions

The stability of mutualistic networks is promoted by a highly connected and nested architecture, whereas stability in antagonistic networks is promoted by a compartmentalized and weakly connected structure (Morris et al. 2014). The subnetworks with the highest number of compartments were those based on Curculionidae, Fabaceae and functional group B (large dry fruits, protected and dehiscent) at BCI. Webs that are strongly compartmentalised (i.e. have high modularity)

might be expected to be stable (across modules) because changes in abundance (or extinction) of individual species (within modules) are less likely to cascade to affect nodes in other parts of the network beyond the affected compartment or module (Thébault and Fontaine 2010). In antagonistic insect-plant networks where the host is immobile (a property that distinguishes them from many other food webs), modularity will often result from trait matching and phylogenetic conservatism in plant traits. High levels of trait matching in most cases will make insect herbivores particularly prone to co-extinction following loss of their host plants. Interactions such as those for Stratiomyidae at Wanang which display lower trait matching may be more robust to random plant species loss but the subnetwork overall will be less resilient to the loss of key nodes rich in fleshy fruit (e.g. well-connected plant genera).

Seed dispersal networks have on average a low specialization (H2') compared to our seed predation subnetworks (Blüthgen et al. 2007: average 0.28; average 0.79 for all our subnetworks). Low H2' promotes high redundancy and increased seed dispersal (Blüthgen et al. 2007). Everything else being equal, plant species supporting generalist dispersers but specialized seed predators with low attack rates may be able to produce large number of viable seeds and may be at an advantage over other plant competitors. This is in line with the plant defence syndrome of high nutritional quality and defence (Agrawal and Fishbein 2006). Further, reviewing seed dispersal networks in the tropics, Escribano-Avila et al. (2018) indicated that woody plants bearing small juicy berries containing many tiny seeds often represent keystone species. From the viewpoint of conserving insects feeding on fruits/seeds, the plants most important in seed predation networks are those which support

many insect species (i.e. with high number of consumer species per host plant species, vulnerability). Although we did not study many of the plant families considered by Escribano-Avila et al. (2018), we note that our plant families with high vulnerability (Supporting information) usually do not bear berries with tiny seeds. This indicates that, from a conservation viewpoint, key plant species in the tropics may differ between networks of seed dispersal and seed predation.

In summary, our study emphasizes the duality between seed dispersal and seed predation networks in the tropics as the former are not very specific whereas the latter are far more specialized and may include different key plant species. From the viewpoint of forest regeneration and conservation, this underlines the need to study both types of network including a variety of potential key plant species.

Acknowledgements – This article is dedicated to the late Larry J. Orsak, who was an inspiration for many of us and promoted actively the work of parabiologists and forest conservation in Papua New Guinea. We thank ForestGEO and the Smithsonian Tropical Research Institute (Panama), Khao Chong Botanical Garden (Thailand) and New Guinea Binatang Research Centre and Wanang Conservation Area (Papua New Guinea) for logistical support. D. Catalina Fernandez, Indira Simon Chaves, Marjorie Cedeño, Marleny Rivera (Panama), Pitoon Kongnoo, Montarika Panmeng, Sutipun Putnaul (Thailand), Dominic Rinan, Jonah Philips, Roll Lilip (Papua New Guinea) collected most of the insect material. Colleagues acknowledged in Basset et al. (2018) helped with the identification of the material.

Funding - This study was supported by ForestGEO and the Czech Science Foundation (GAČR 20-31295S to YB). Field work on BCI was largely funded by a postdoctoral grant from the Academy of Finland to SG. Grants from the Smithsonian Institution Barcoding Opportunity FY013 and FY014 (to YB), from the ForestGEO Research Grant Program (to CD), and in-kind help from the Canadian Centre for DNA Barcoding and Southern China DNA Barcoding Center allowed to sequence insect specimens. YB and HB were supported by the Sistema Nacional de Investigación, SENACYT, Panama. SG holds a Royal Society Univ. Research Fellowship. MK was partly supported by the Ministry of Agriculture of the Czech Republic, institutional support MZE-RO0118. DLJQ was supported by a Senior Postdoctoral Fellowship under Ratchadaphiseksomphot Fund, Graduate School, Chulalongkorn Univ. BAB was funded by Ratchadaphiseksomphot Endowment Fund Chulalongkorn Univ. (R/F_2559_019_05_23). VN was supported by European Research Council grant 669609. STS acknowledges Deptal support from Harper Adams Univ.

Conflicts of interest – The authors have declared that no conflicts of interest exist.

Permits – Research permits were delivered by the Smithsonian Tropical Research Institute (BCI), the National Research Council of Thailand (0002/825; KHC) and the Conservation and Environment Protection Agency (15233, 16091, 16090, 16093, 16207; WAN).

Author contributions

Yves Basset: Conceptualization (lead); Data curation (lead); Formal analysis (lead); Funding acquisition (equal); Project administration (lead); Writing – original draft (lead); Writing

- review and editing (lead). **Leonardo R. Jorge**: Formal analysis (equal); Writing – original draft (equal); Writing – review and editing (equal). Philip T. Butterill: Formal analysis (equal); Writing – original draft (equal); Writing – review and editing (equal). Greg P. A. Lamarre: Formal analysis (equal); Writing – original draft (equal); Writing – review and editing (equal). Chris Dahl: Conceptualization (equal); Data curation (equal); Funding acquisition (equal); Writing - original draft (equal); Writing - review and editing (equal). Richard Ctvrtecka: Conceptualization (equal); Data curation (equal); Writing – original draft (equal); Writing – review and editing (equal). Sofia Gripenberg: Conceptualization (equal); Data curation (equal); Writing - original draft (equal); Writing review and editing (equal). **Owen T. Lewis**: Conceptualization (equal); Writing – original draft (equal); Writing – review and editing – (equal). **Héctor Barrios**: Conceptualization (equal); Data curation (equal); Writing - original draft (equal); Writing – review and editing (equal). John W. Brown: Data curation (equal); Writing – original draft (equal); Writing – review and editing (equal). Sarayudh Bunyavejchewin: Data curation (equal); Writing - original draft (equal); Writing review and editing (equal). Buntika A. Butcher: Data curation (equal); Writing – original draft (equal); Writing – review and editing (equal). Anthony I. Cognato: Data curation (equal); Writing - original draft (equal); Writing - review and editing (equal). Stuart J. Davies: Data curation (equal); Writing – original draft (equal); Writing – review and editing (equal). Ondrej Kaman: Data curation (equal); Writing original draft (equal); Writing – review and editing (equal). Petr Klimeš: Data curation (equal); Writing – original draft (equal); Writing – review and editing (equal). Milos Knížek: Data curation (equal); Writing - original draft (equal); Writing – review and editing (equal). Scott E. Miller: Data curation (equal); Writing - original draft (equal); Writing review and editing (equal). Geoffrey E. Morse: Data curation (equal); Writing - original draft (equal); Writing - review and editing (equal). Vojtech Novotny: Data curation (equal); Writing – original draft (equal); Writing – review and editing (equal). Nantachai Pongpattananurak: Data curation (equal); Writing - original draft (equal); Writing - review and editing (equal). Pairot Pramual: Data curation (equal); Writing - original draft (equal); Writing - review and editing (equal). **Donald L. J. Quicke**: Data curation (equal); Writing - original draft (equal); Writing - review and editing (equal). Watana Sakchoowong: Data curation (equal); Writing – original draft (equal); Writing – review and editing (equal). Ruma Umari: Data curation (equal); Writing original draft (equal); Writing – review and editing (equal). **Eero J. Vesterinen**: Data curation (equal); Writing – original draft (equal); Writing - review and editing (equal). George Weiblen: Data curation (equal); Writing – original draft (equal); Writing - review and editing (equal). S. Joseph Wright: Data curation (equal); Formal analysis (equal); Writing – original draft (equal); Writing – review and editing (equal). **Simon T. Segar**: Data curation (equal); Formal analysis (equal); Writing - original draft (equal), Writing - review and editing (equal).

Data availability statement

Data available from Figshare Digital Repository: http://doi.org/10.25573/data.11444571.v1 (Basset et al. 2021).

References

- Agrawal, A. A. and Fishbein, M. 2006. Plant defense syndromes. Ecology 87: S132–S149.
- Anderson-Teixeira, K. J. et al. 2014. CTFS-ForestGEO: a world-wide network monitoring forests in an era of global change. Global Change Biol. 21: 528–549.
- Basset, Y. 2001. Communities of insect herbivores foraging on mature trees vs seedlings of *Pourouma bicolor* (Cecropiaceae) in Panama. – Oecologia 129: 253–260.
- Basset, Y. et al. 2018. A cross-continental comparison of assemblages of seed- and fruit-feeding insects in tropical rainforests: faunal composition and rates of attack. J. Biogeogr. 45: 1395–1407.
- Basset, Y. et al. 2021. Data from: Host specificity and interaction networks of insects feeding on seeds and fruits in tropical rainforests. Figshare Digital Repository, http://doi.org/10.25573/data.11444571.v1.
- Bazzaz, F. A. et al. 1987. Allocating resources to reproduction and defense. BioScience 37: 58–67.
- Berenbaum, M. R. 2001. Chemical mediation of coevolution: phylogenetic evidence for Apiaceae and associates. Ann. Miss. Bot. Gard. 88: 45–59.
- Blüthgen, N. et al. 2006. Measuring specialization in species interaction networks. BMC Ecol. 6: 9.
- Blüthgen, N. et al. 2007. Specialization, constraints and conflicting interests in mutualistic networks. Curr. Biol. 17: 341–346.
- Chamberlain, S. A. et al. 2014. Traits and phylogenetic history contribute to network structure across Canadian plant–pollinator communities. Oecologia 176: 545–556.
- Chase, M. W. et al. 2016. An update of the Angiosperm Phylogeny Group classification for the orders and families of flowering plants: APG IV. – Bot. J. Linn. Soc. 181: 1–20.
- Copeland, R. S. et al. 2009. Insects reared from the wild fruits of Kenya. J. E. Afr. Nat. Hist. 98: 11–66.
- Ctvrtecka, R. et al. 2014. Frugivorous weevils are too rare to cause Janzen–Connell effects in New Guinea lowland rain forest. J. Trop. Ecol. 30: 521–535.
- Ctvrtecka, R. et al. 2016. Fruit sizes and the structure of frugivorous communities in a New Guinea lowland rainforest. Austr. Ecol. 43: 228–237.
- Dahl, C. et al. 2019. The insect-focused classification of fruit syndromes in tropical rainforests: an inter-continental comparison.

 Biotropica 51: 39–49.
- Dáttilo, W. and Rico-Gray, V. 2018. Ecological networks in the tropics: an integrative overview of species interactions from some of the most species-rich habitats on Earth. – Springer.
- de Aguiar, M. A. et al. 2019. Revealing biases in the sampling of ecological interaction networks. PeerJ 7: e7566.
- Dormann, C. F. et al. 2009. Indices, graphs and null models: analyzing bipartite ecological networks. Open Ecol. J. 2: 7–24.
- Dormann, C. F. et al. 2018. Package 'bipartite'. ver. 2.11. https://github.com/biometry/bipartite.
- Ehrlich, P. R. and Raven, P. H. 1964. Butterflies and plants: a study in coevolution. Evolution 18: 586–608.
- Escribano-Avila, G. et al. 2018. Tropical seed dispersal networks: emerging patterns, biases and keystone species traits. In: Dát-

- tilo, W. and Rico-Gray, V. (eds), Ecological networks in the tropics. Springer, pp. 93–110.
- Feeny, P. P. 1976. Plant apparency and chemical defense. Recent Adv. Phytochem. 10: 1–40.
- Fox, J. and Weisberg, S. 2019. An R companion to applied regression, 3rd edn. Sage, Thousand Oaks, CA.
- Gauld, I. D. et al. 1992. Plant allelochemicals, tritrophic interactions and the anomalous diversity of tropical parasitoids: the 'nasty' host hypothesis. Oikos 65: 353–357.
- Gripenberg, S. 2018. Do pre-dispersal insect seed predators contribute to maintaining tropical forest plant diversity? Biotropica 50: 839–845.
- Gripenberg, S. et al. 2018. Seed polyphenols in a diverse tropical plant community. J. Ecol. 106: 87–100.
- Gripenberg, S. et al. 2019. A highly resolved food web for insect seed predators in a species-rich tropical forest. Ecol. Lett. 22: 1638–1649.
- Gruen, B., Kosmidis, I. et al. 2012. Extended beta regression in R: shaken, stirred, mixed and partitioned. J. Stat. Softw. 48: 1–25
- Hawkins, B. A. and Lawton, J. H. 1987. Species richness for parasitoids of British phytophagous insects. – Nature 326: 788–790.
- Hopkins, M. J. G. 1983. Unusual diversities of seed beetles (Coleoptera: Bruchidae) on *Parkia* (Leguminosae: Mimosoideae) in Brazil. Biol. J. Linn. Soc. 19: 329–338.
- Ives, A. R. and Godfray, H. C. J. 2006. Phylogenetic analysis of trophic associations. Am. Nat. 168: 1, E1–E14.
- Janzen, D. H. 1971. Seed predation by animals. Annu. Rev. Ecol. Syst. 2: 465–492.
- Janzen, D. H. 1976. Seeding patterns of tropical trees. In: Tomlinson, P. B. and Zimmermann, M. H. (eds), Tropical trees as living systems. Cambridge Univ. Press, pp. 88–128.
- Janzen, D. H. 1980. Specificity of seed-attacking beetles in a Costa Rican deciduous forest. – J. Ecol. 68: 929–952.
- Jordano, P. 2016. Sampling networks of ecological interactions. Funct. Ecol. 30: 1883–1893.
- Jordano, P. et al. 2003. Invariant properties in coevolutionary networks of plant–animal interactions. Ecol. Lett. 6: 69–81.
- Jorge, L. R. et al. 2014. An integrated framework to improve the concept of resource specialisation. Ecol. Lett. 17: 1341–1350.
- Jorge, L. R. et al. 2017. Phylogenetic trophic specialization: a robust comparison of herbivorous guilds. Oecologia 187: 551–559.
- Kembel, S. W. et al. 2010. Picante: R tools for integrating phylogenies and ecology. Bioinformatics 26: 1463–1464.
- Kergoat, G. J. et al. 2005. Both host-plant phylogeny and chemistry have shaped the African seed-beetle radiation. Mol. Phylogenet. Evol. 35: 602–611.
- Kissling, W. D. et al. 2009. The global distribution of frugivory in birds. Global Ecol. Biogeogr. 18: 150–162.
- Laliberté, E. and Legendre, P. 2010. A distance-based framework for measuring functional diversity from multiple traits. Ecology 91: 299–305.
- Laliberté, E. et al. 2014. FD: measuring functional diversity from multiple traits, and other tools for functional ecology. R package ver. 1.0-12, https://rdrr.io/cran/FD/>.
- Lewinsohn, T. M. and Roslin, T. 2008. Four ways towards tropical herbivore megadiversity. Ecol. Lett. 11: 398–416.
- Lewinsohn, T. M. et al. 2006. Structure in plant–animal interaction assemblages. Oikos 113: 174–184.
- Lewis, O. T. and Gripenberg, S. 2008. Insect seed predators and environmental change. J. Appl. Ecol. 45: 1593–1599.

- Morris, R. J. et al. 2014. Antagonistic interaction networks are structured independently of latitude and host guild. Ecol. Lett. 17: 340–349.
- Nakagawa, M. et al. 2003. Resource use of insect seed predators during general flowering and seeding events in a Bornean dipterocarp rain forest. Bull. Entomol. Res. 93: 455–466.
- Novotny, V. and Basset, Y. 2005. Host specificity of insect herbivores in tropical forests. Proc. R. Soc. B 272: 1083–1090.
- Novotny, V. et al. 2002. Low host specificity of herbivorous insects in a tropical forest. Nature 416: 841–844.
- Novotny, V. et al. 2010. Guild-specific patterns of species richness and host specialization in plant–herbivore food webs from a tropical forest. J. Anim. Ecol. 79: 1193–203.
- Olesen, J. M. and Jordano, P. 2002. Geographic patterns in plant–pollinator mutualistic networks. Ecology 83: 2416–2424.
- Paniagua, M. R. et al. 2009. Structure and vertical stratification of plant galler–parasitoid food webs in two tropical forests. – Ecol. Entomol. 34: 310–320.
- Poisot, T. et al. 2015. Beyond species: why ecological interaction networks vary through space and time. Oikos 124: 243–251.
- Prado, P. I. and Lewinsohn, T. M. 2004. Compartments in insect– plant associations and their consequences for community structure. – J. Anim. Ecol. 73: 1168–1178.
- Price, P. W. 1992. The resource-based organization of communities. Biotropica 24: 273–282.
- Qian, H. and Jin, Y. 2016. An updated megaphylogeny of plants, a tool for generating plant phylogenies and an analysis of phylogenetic community structure. J. Plant Ecol. 9: 233–239.
- Quicke, D. L. 2012. We know too little about parasitoid wasp distributions to draw any conclusions about latitudinal trends in species richness, body size and biology. PLoS One 7: e32101.
- Quicke, D. L. J. 2015. The braconid and ichneumonid parasitoid wasps: biology, systematics, evolution and ecology. Wiley.
- Quinto, J. et al. 2012. Breaking down complex saproxylic communities: understanding sub-networks structure and implications to network robustness. PLoS One 7: e45062.
- Redmond, C. M. et al. 2018. High specialization and limited structural change in plant–herbivore networks along a successional chronosequence in tropical montane forest. Ecography 42: 162–172.
- Rønsted, N. et al. 2012. Can phylogeny predict chemical diversity and potential medicinal activity of plants? A case study of Amaryllidaceae. – BMC Evol. Biol. 12: 182.

- Root, R. B. 1973. Organization of a plant–arthropod association in simple and diverse habitats: the fauna of collards *Brassica oleracea*. Ecol. Monogr. 43: 95–124.
- Sam, K. et al. 2017. Low host specificity and abundance of frugivorous Lepidoptera in the lowland rain forests of Papua New Guinea. PLoS One 12: e0171843.
- Schleuning, M. et al. 2012. Specialization of mutualistic interaction networks decreases toward tropical latitudes. Curr. Biol. 22: 1925–1931.
- Sedio, B. E. et al. 2018. Comparative foliar metabolomics of a tropical and a temperate forest community. Ecology 99: 2647–2653.
- Segar, S. T. et al. 2017. Varyingly hungry caterpillars: predictive models and foliar chemistry suggest how to eat a rainforest. – Proc. R. Soc. B 284: 20171803.
- Segar, S. T. et al. 2020. The role of evolution in shaping ecological networks. Trends Ecol. Evol. 35: 454–466.
- Smilanich, A. M. et al. 2009. Immunological cost of chemical defence and the evolution of herbivore diet breadth. – Ecol. Lett. 12: 612–621.
- Stephenson, A. G. 1981. Flower and fruit abortion: proximate causes and ultimate functions. Annu. Rev. Ecol. Syst. 12: 253–279.
- Thébault, E. and Fontaine, C. 2010. Stability of ecological communities and the architecture of mutualistic and trophic networks. Science 329: 853–856.
- Volf, M. et al. 2017. Community structure of insect herbivores is driven by conservatism, escalation and divergence of defensive traits in *Ficus*. – Ecol. Lett. 21: 83–92.
- von Oertzen, T. et al. 2015. Structural equation modeling with Ωnyx. Struct. Equat. Model. 22: 148–161.
- Webb, C. O. 2000. Exploring the phylogenetic structure of ecological communities: an example for rain forest trees. Am. Nat. 156: 145–155.
- Weiblen, G. D. et al. 2006. Phylogenetic dispersion of host use in a tropical insect herbivore community. Ecology 87: S62–S75.
- Zangerl, A. R. and Bazzaz, F. A. 1992. Theory and pattern in plant defense allocation. In: Fritz, R. S. and Simms, E. L. (eds), Plant resistance to herbivores and pathogens: ecology, evolution and genetics. Univ. of Chicago Press, pp. 363–391.
- Zanne, Å. E. et al. 2014. Three keys to the radiation of angiosperms into freezing environments. Nature 506: 89–92.